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Short communication

Simultaneous direct high-performance liquid chromatographic enantioseparation of 2-methylglycidol-1-benzyl ether and 2methylglycerol-1-benzyl ether using a solvent-switching technique

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Abstract

Simultaneous HPLC separation of the enantiomers of 3-benzyloxy-2-methyl-1,2-propanediol and the corresponding 3-benzyloxy-2-methyl-1,2-propene oxide could be accomplished on amylose derived Chiralpak AD switching between 10% 2-propanol and 3% 1,2-dimethoxyethane as polar modifier in *n*-heptane. © 2001 Elsevier Science B.V. All rights reserved.

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1. Introduction

Carbaldehyde **1**, which is an important building block [1], can be prepared starting from mannitol [2]. This approach has the disadvantage of leading only to the (R)-enantiomer (six steps, total yield 13%). A biocatalytic kinetic resolution by lipase led to high enantiomeric excess (ee>95%) but again only with low yield [3]. Sharpless epoxidation of 2-methyl-3propenol [4] and dihydroxylation of 3-benzyloxy-2methylpropene [5] lead to products with rather low enantiomeric excesses (ee=85% and 77%, respectively). In the course of our investigation of an enantioconvergent chemo-enzymatic way to prepare

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(S)-1 [1] we decided to investigate the enzymatic resolution of O-benzyl protected 2-methylglycidol 2, which can be easily converted to 1 (Fig. 1). Protected hydroxymethyl epoxides are common substrates in biotransformation steps [6]. The development of an analytical method to determine the enantiomeric excess of epoxide 2 and diol 3 posed a problem. Previous successfully employed gas chromatographic methods to analyze chiral analogues containing a secondary carbon in position 2 [6] or tertiary substituted analogues containing a longer aliphatic chain [7] failed in the case of both benzyl ethers 2 and 3. The gas-liquid chromatographic separation of the enantiomers of simple glycidol and glycerol analogues 4 and 5 (the latter derivatized as carbonate or acetonide) is known [8] but high-performance liquid chromatographic separation of 2 and 3 has not been described in the literature. Separation of the enantiomers of 3-tosyloxy-2-methylpropene

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Fig. 1. Structures of carbaldehyde $\mathbf{1}$ and the investigated benzyl ethers.

oxide [9] and 2-methylglycerol as 1-(4-methoxybenzoate) [10] has been reported.

In this paper we describe our results using gasliquid chromatography (GLC) and high-performance liquid chromatography (HPLC) chiral stationary phases and the optimized simultaneous liquid chromatographic resolution of **2** and **3** on amylose derived Chiralpak AD.

2. Experimental

2.1. Chemicals and reagents

Synthesis of 1 [2], 2, 3 and 3a [6] has been reported.

Compound **4** was obtained from Sigma–Aldrich (Vienna, Austria). *n*-Heptane, 2-propanol (for liquid chromatography) and 1,2-dimethoxyethane were obtained from Merck (Darmstadt, Germany).

2.2. Chromatography

GLC measurements were carried out on a Varian 3800 gas chromatograph equipped with a flame ionization detection (FID) system using H₂ as the carrier gas. Investigated columns were: Chrompak Chirasil-dex CB (β -cyclodextrin directly bonded to a dimethylpolysiloxane; 25 m×0.32 mm I.D., 0.25 µm film), ASTEC Chiraldex B-TA column (β -cyclodextrin, modified as 2,6-di-*O*-pentyl-3-trifluoroacetyl derivative; 30 m×0.25 mm I.D.), ASTEC Chiraldex G-PN (γ -cyclodextrin as the 2,6-di-*O*-pentyl-3-propionyl derivative; 30 m×0.25 mm I.D.), ASTEC Chiraldex G-PH [γ -cyclodextrin as (*S*)-2-hydroxy-propyl methyl ether; 30 m×0.25 mm I.D.].

HPLC analyses were performed on a Jasco system containing a PU-980 pump equipped with an MD-910 multiwavelength detector. Investigated columns were: Daicel Chiralpak AD ($250 \times 4.6 \text{ mm}$), Daicel Chiralcel ODH ($250 \times 4.6 \text{ mm}$), Merck (3S,4R) Whelk-O1 ($250 \times 4 \text{ mm}$).

Hold-up times were determined using 1,3,5-tri*tert.*-butylbenzene. Separation parameters for **3** were calculated as started from the moment of solvent switch in order to obtain relevant numbers.

3. Results and discussion

3.1. GLC approaches

Racemic secondary carbon containing compounds 4 and 5 could be separated via gas chromatography using literature conditions [8]. However, tertiary analogues 2 and 3 resisted several attempts on all four investigated columns (see Experimental). We could separate the enantiomers as 1-methoxy derivative 3a on Chirasil-Dex [6], but the method was not practical due to many side products.

3.2. HPLC approaches

Attempts failed to separate **2** and **3** on Pirkle-type CSP Whelk-O1, which has shown previously enantioseparation abilities for compounds with these types of functional groups. [**2**: $k'_2=0.65$; **3**: $k'_2=1.05$; conditions: *n*-heptane-2-propanol (90:10), 0.5 ml/min, 20°C]. Better results were obtained using

А

0

В

10

20

cellulose 3,5-dimethylphenylcarbamate derived Chiralcel ODH. Diol **3** was partly separated $[k'_2=1.61, \alpha=1.08, R_s=0.88;$ conditions: *n*-heptane–2-propanol (90:10), 0.5 ml/min, 20°C], epoxide **2** remained unresolved $[k'_2=0.36;$ conditions: *n*-heptane– 2-propanol (95:5), 0.5 ml/min, 20°C]. Use of less 2-propanol as polar modifier did not improve results.

Amylose 3,5-dimethylphenylcarbamate derived Chiralpak AD showed the relatively best performance: optimized flow and temperature conditions for diol **3** resulted in baseline separation $[k'_2=2.22, \alpha=$ 1.12, $R_s=1.85$; conditions: *n*-heptane-2-propanol (90:10), 0.5 ml/min, 20°C]. However, epoxide **2** remained poorly separated [Fig. 2A; $k'_2=0.96$, $\alpha=$ 1.07, $R_s=0.3$; conditions: *n*-heptane-2-propanol (98:2), 0.5 ml/min, 10°C].

Changing the polar modifier from routinely used 2-propanol to aprotic 1,2-dimethoxyethane (mono-

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glyme), far better results for epoxide **2** could be obtained, especially after careful optimization of the temperature [Fig. 2B; k'_2 =3.50, α =1.14, R_s =1.68; conditions: *n*-heptane-monoglyme (98:2), 0.5 ml/min, 10°C].

In screening procedures it is highly desirable to observe both, educt and product simultaneously. This could be accomplished by using a solvent switching method starting with the less polar mobile phase containing monoglyme resolving **2** (k'_2 =2.49, α = 1.13, R_s =1.51) and leaving the chiral diol **3** at the head of the column. Changing the mobile phase modifier to 2-propanol and optimizing flow-rates it was possible to elute baseline separated enantiomers of diol **3** [k'_2 =1.74, α =1.16, R_s =1.31; conditions: 10°C; 0-35 min: *n*-heptane–monoglyme (97:3), 0.5 ml/min, 35-45 min: *n*-heptane–2-propanol (90:10), 0.75 ml/min, 45-80 min: *n*-heptane–2-propanol

switch

40

30

min

OCH,C

60

CH_C_H

(R

50



 $\begin{array}{c|c} & & & & & & & \\ & & & & & \\$

Fig. 2. Effects on the separation of racemic epoxide 2 using 2% 2-propanol (A) or monoglyme (B) as polar modifier in *n*-heptane (Chiralpak AD, conditions see Results and discussion).

Fig. 3. Separation of epoxide **2** and diol **3** on Chiralpak AD using optimized conditions (A) and slightly different conditions (B) (see Results and discussion).

(90:10), 0.5 ml/min, Fig. 3A]. We can show in Fig. 3B, that small changes of the optimized protocol give far less useful results [**2**: k'_2 =1.27, α =1.09, R_s =0.88; **3**: k'_2 =0.89, α =1.12, R_s =1.13; conditions: 10°C, 0-35 min: *n*-heptane–monoglyme (96:4), 0.5 ml/min, 35-55 min: *n*-heptane–2-propanol (90:10), 0.75 ml/min, 55-80 min: *n*-heptane–2-propanol (90:10), 0.5 ml/min].

4. Conclusions

We have shown, that carefully chosen separation conditions in combination with a solvent switching technique lead to a simultaneous HPLC enantio-separation of 2-methylglycidol-1-benzyl ether 2 and 2-methylglycerol-1-benzyl ether 3 facilitating enzymatic hydrolysis studies.

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